

BACKGROUND

Phospholipase C (PLC) enzymes, comprising several families (PLC β , γ , δ , ϵ , η , and ζ), have been established as crucial signaling molecules involved in regulation of a variety of cellular functions. PLC-catalyzed formation of the second messengers, inositol 1,4,5-trisphosphate (IP₃) and diacylglycerol, from phosphatidylinositol 4,5-bisphosphate (PIP₂), constitutes one of the major cell signaling responses. IP₃ induces a transient increase in intracellular free Ca²⁺, while DAG directly activates protein kinase C. These second messengers provide a common link from highly specific receptors for hormones, neurotransmitters, antigens, and growth factors to downstream, intracellular targets; thus, they contribute to regulation of biological functions as diverse as cell motility, fertilization, and sensory transduction.¹

Of two PLC γ enzymes, PLC γ 1 is ubiquitously expressed and appears to regulate a multitude of cellular functions in many tissues. It is activated in response to growth factor stimulation; in addition, its function in T-cell responses has been extensively documented. PLC γ 2, in contrast, is most highly expressed in cells of the hematopoietic system and plays a key role in regulation of the immune response. Both PLC γ enzymes have also been implicated in signaling events underlying aberrant cellular responses. PLC γ 1 is critically involved in the regulation of cancer cell motility while PLC γ 2 has been implicated in deregulation of the immune responses resembling Btk-dependent X-linked agammaglobulinemia and SLE disease in humans. It has been suggested that, in cancer cells, PLC γ 1 could function as a key, rate-limiting, common component involved in cell motility triggered by several growth factors and integrins.²

The domain organization of PLC γ enzymes is characterized by the insertion of a highly structured region (PLC γ -specific array, gammaSA) between the two halves of the TIM-barrel catalytic domain common to all PLCs. The gammaSA comprises a split PH (spPH) domain flanking two tandem SH2 domains and a SH3 domain. A distinct regulatory feature of PLC γ enzymes is that their activation is linked to an increase in phosphorylation of specific tyrosine residues (most notably within the gammaSA) by receptor and non-receptor tyrosine kinases. Upon stimulation of cells with PDGF and EGF, the SH2 domain of PLC- γ binds to the autophosphorylated tyrosine residues of growth factor receptors, leading to tyrosine phosphorylation and activation of PLC- γ .³ Full activation of PLC gamma 2 requires phosphorylation of Tyr753, 759, 1197, and 1217. All four tyrosines are phosphorylated following BCR engagement, most likely by Btk.⁴ In addition, activation of PLC- γ isozymes may occur secondarily to receptor-mediated activation of

phospholipase D and cytosolic phospholipase A₂, which results in the production of phosphatidic acid and arachidonic acid, respectively.⁵ It was also reported that PLC- γ is regulated additionally by the lipid products of PI 3-kinase. The PH domain of PLC- γ binds to PtdIns(3,4,5)P₃, and is targeted to the membrane in response to growth factor stimulation and leads to activation of PI 3-kinase causes PLC- γ PH domain-mediated membrane targeting and PLC- γ activation.⁶ Furthermore, multiple protein-protein interactions (mainly mediated by SH2 domains) also contribute to activation and have an important role in localizing PLC γ into protein complexes with different binding partners, depending on cell type and specific cellular compartments. One mode of activation that is specific for the PLC γ 2 isozyme is direct binding to and activation by Rac. The interaction involves the spPH domain, and this activation mechanism does not require tyrosine phosphorylation.⁷

References:

1. Kim, M.G. et al: *Exp. Mol. Med.* 32:101-9, 2000
2. Patterson, R.L. et al: *Trends in Biochem. Chem.* 30:688-97, 2005
3. Nishibe, S. & Carpenter, G.: *Semin Cancer Biol.* 1:285-92, 1990
4. Watanabe, D. et al: *J. Biol. Chem.* 276:38595-601, 2001
5. Sekiya, F. et al: *Chem. Phys. Lipids* 98:3-11, 1998
6. Falasca, M. et al: *EMBO J.* 17:414-22, 1998
7. Carpenter, G. & Ji, Q.: *Exp. Cell Res.* 253:15-24, 1999

TECHNICAL INFORMATION

Source:

Phospho-PLC gamma 2 (Tyr1217) antibody is a rabbit antibody raised against a short peptide from human PLC gamma 2 sequence surrounding and containing phospho-Tyr1217.

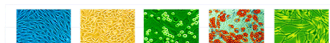
Specificity and Sensitivity:

This antibody detects endogenous phosphorylated PLC gamma 2 (Tyr1217) proteins without cross-reactivity with other family members.

Storage Buffer: Rabbit IgG in phosphate buffered saline (without Mg²⁺ and Ca²⁺), pH 7.4, 150mM NaCl, 0.02% sodium azide and 50% glycerol.

Storage:

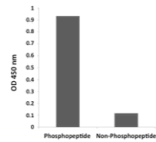
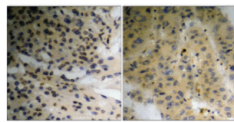
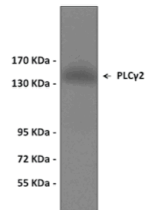
Store at -20°C for at least one year. Store at 4°C for frequent use. Avoid repeated freeze-thaw cycles.



APPLICATIONS

Application:	*Dilution:
WB	1:500-1:1000
IP	n/d
IHC	1:50-1:100
ICC	n/d
FACS	n/d
ELISA	1:1000

QUALITY CONTROL DATA



Top: Western blot analysis of extracts from Jurkat cells treated with UV for 5 minutes.

Middle: Immunohistochemistry analysis of paraffin-embedded Human brain gliomas (left) and Human liver carcinoma (right).

Bottom: ELISA for Immunogen Phosphopeptide (left) and Non-Phosphopeptide (right).

